

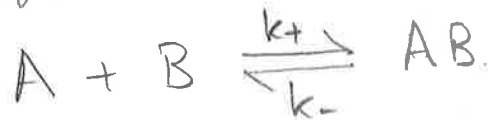
Lecture 3 Hemoglobin

1. Law of Mass Action

Can derive formally from the condition for equilibrium $\Delta G = 0$ (p268.)

Include the concentration dependence of the chemical potentials $\mu = \mu_0 + k_B T \ln c/c_0$.

But the result can be seen directly from rate equations:



$$\left. \begin{aligned} \text{Forward rate} &= [A][B]k_+ \\ \text{Reverse rate} &= [AB]k_- \end{aligned} \right\} \text{equal in } \equiv m$$

$$[A][B]k_+ = [AB]k_-$$

$$\Rightarrow \frac{[AB]}{[A][B]} = \frac{k_+}{k_-} = K_{eq} \text{ equilibrium constant.}$$

All chemical reactions can be characterized by an equilibrium constant. Units depend on the "order" of the reaction, eg valency of the reacting species. Here $[K_{eq}] = \text{conc}^{-1}$.

2. Ligand - Receptor binding.

We already saw the "Langmuir isotherm" for binding probability of a ligand to receptor

concentrations: $[L]$ $[R]$



$$\frac{[L][R]}{[LR]} = K_d \text{ equilib. const. (units = conc)}$$

$$P_{\text{bound}} = \frac{[LR]}{[LR] + [R]} = \frac{[LR]/[R]}{1 + [L]/[R]} = \frac{[L]/K_d}{1 + [L]/K_d}$$

This is the same as the result before:

$$P_{\text{bound}} = \frac{(c/c_0) e^{-\beta \Delta \epsilon}}{1 + (c/c_0) e^{-\beta \Delta \epsilon}} \quad \text{where } c = [L]$$

$$\text{Comparing: } K_d = c_0 e^{+\beta \Delta \epsilon}$$

Where c_0 = concentration of lattice sites in solution

$$\Delta \epsilon = \text{binding energy} = \epsilon_b - \epsilon_{\text{sol}} \quad [2.10]$$

Two results allow conversion back/forth from microscopic state params to macro concentrations.

3. Entropy of binding.

Intuitively, there is no entropy associated with $P_{\text{bound}} = 0$ or $P_{\text{bound}} = 1$, fully specified.

Use Shannon Entropy to look in between:

$$S = -\sum_i p_i \ln p_i = \text{Missing Information}$$

Here there are 2 states bound & unbound.

$$S = -P_{\text{bound}} \ln P_{\text{bound}} - P_{\text{unbound}} \ln P_{\text{unbound}}$$

$$\text{with } P_{\text{bound}} + P_{\text{unbound}} = 1$$

$$P_{\text{bound}} = \frac{[L]/K_d}{1 + [L]/K_d} = \frac{\alpha}{1 + \alpha} \quad P_{\text{unbound}} = \frac{1}{1 + \alpha}$$

$$S = -\frac{\alpha}{1 + \alpha} \ln \frac{\alpha}{1 + \alpha} - \frac{1}{1 + \alpha} \ln \frac{1}{1 + \alpha}$$

$$= -\frac{1}{1 + \alpha} [\alpha \ln \alpha - \alpha \ln (1 + \alpha) + \ln (1 + \alpha)]$$

$$= \frac{+1}{1 + \alpha} [(1 + \alpha) \ln (1 + \alpha) - \alpha \ln \alpha]$$

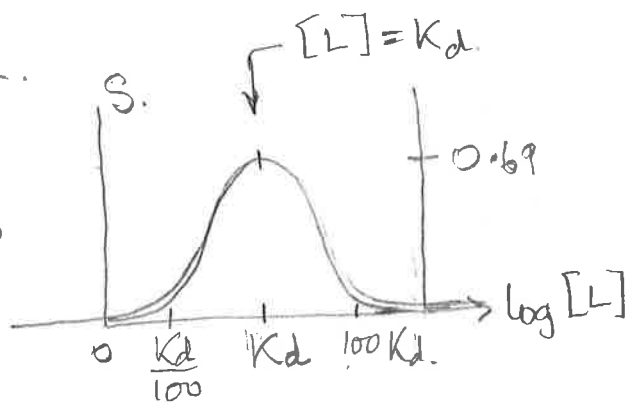
$$= \ln (1 + \alpha) - \frac{\alpha}{1 + \alpha} \ln \alpha$$

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$$\alpha \ll 1 \quad S \rightarrow \alpha - \alpha \ln \alpha \rightarrow 0$$

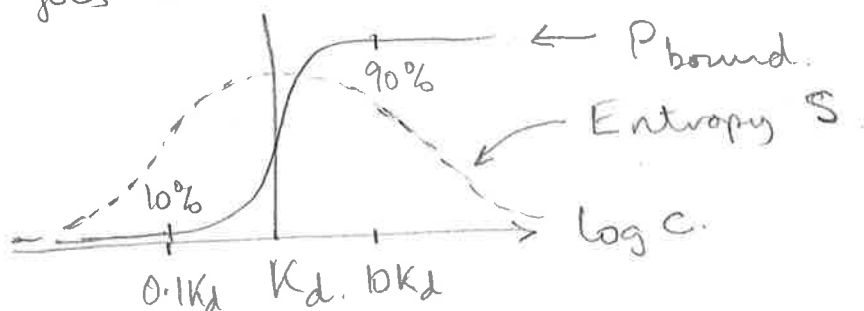
$$\alpha \gg 1 \quad S \rightarrow \ln \alpha - \ln \alpha \rightarrow 0$$

$$\alpha = 1 \quad S = \ln 2 - 0 \neq 0$$



3.3

This goes with the "titration curve" we saw before



$\log c$ is the same scale as energy or chem potential
 or pH for $[H^+]$

$$\mu = \mu_0 + k_B T \ln c/c_0.$$

4. Hill Function

Treat higher order reactions the same way.
 When two ligands bind the receptor at the same time:



$$\left. \begin{aligned} \text{Forward rate} &= [L]^2 [R] k_+ \\ \text{Backward} &= [L_2R] k_- \end{aligned} \right\} K_d^2 = \frac{[L]^2 [R]}{[L_2R]}$$

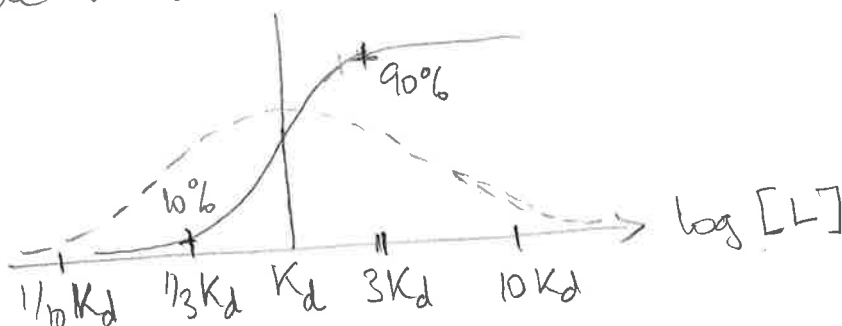
New K_d is squared to keep units of conc.

$$P_{\text{bound}} = \frac{[L_2R]}{[L_2R] + [R]} = \frac{([L]/K_d)^2}{1 + ([L]/K_d)^2}$$

Generalise to $n+1$ th-order reaction [n is a measure of cooperativity]

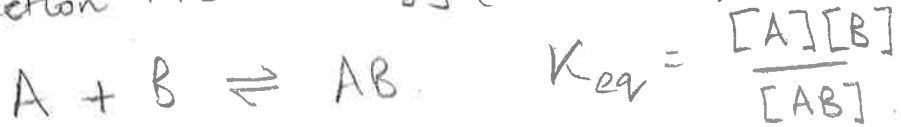
$$P_{\text{bound}} = \frac{([L]/K_d)^n}{1 + ([L]/K_d)^n} \quad \text{Hill function}$$

Shannon entropy, as before, with $\alpha = ([L]/K_d)^n$
 Both curves get narrower, as seen on log scale most easily:



3.4

5. Reaction Free Energy (out of equilib).



Definition $\mu = \frac{\partial G}{\partial N}$ chemical potential.

$$\Delta G = \mu_A \Delta N_A + \mu_B \Delta N_B + \mu_{AB} \Delta N_{AB} \quad \left. \begin{array}{l} \text{one} \\ \text{molecule} \\ \text{reacts} \end{array} \right\}$$

$\begin{matrix} +1 & +1 & -1 \end{matrix}$

Recalling that $\mu_A = \mu_{A0} + k_B T \ln [A] / [\text{ref} A]$

$$\Delta G = \mu_{A0} + k_B T \ln \frac{[A]}{[\text{ref} A]} + \mu_{B0} + k_B T \ln \frac{[B]}{[\text{ref} B]} - \mu_{AB0} - k_B T \ln \frac{[AB]}{[\text{ref} AB]}$$

$$= \mu_{A0} + \mu_{B0} - \mu_{AB0} + k_B T \ln \frac{[A][B]}{[AB]} - k_B T \ln \frac{[\text{ref} A][\text{ref} B]}{[\text{ref} AB]}$$

$$= \Delta G_0 + k_B T \ln \frac{[A][B]}{[AB]}$$

where ΔG_0 includes the equilibrium concs of the species: $\Delta G_0 = -k_B T \ln K_{eq}$.

Note: if reaction is in equilibrium, the concentrations obey K_{eq} def and $\Delta G = 0$.

But out of equilibrium reaction generates a driving force

6. Examples

i) ATP drives many energetic processes in cells



Standard state free energy $\Delta G_0 = -12.5 k_B T / \text{mol}$.

Typical *E. coli* numbers:

$$[ATP] = 5 \text{ mM}$$

$$[ADP] = 0.5 \text{ mM}$$

$$[P_i] = 10 \text{ mM}$$

$$\frac{[ADP][P_i]}{[ATP]} = \frac{0.5 \times 10}{5} \times 10^{-3} = 10^{-3}$$

$$\ln 10^{-3} = -6.9$$

same as
Coca cola.

$$\Delta G = (-12.5 - 6.9) k_B T = -19.4 k_B T \text{ in } E. coli$$

ii) Expts to determine K_{eq} .

Measure some physical property such as light absorption spectrum:

A. Myoglobin changes colour when O_2 binds.



B. HIV protein to CD4 receptor in solution.

C. Transcription factor NtrC to DNA.

Methods:

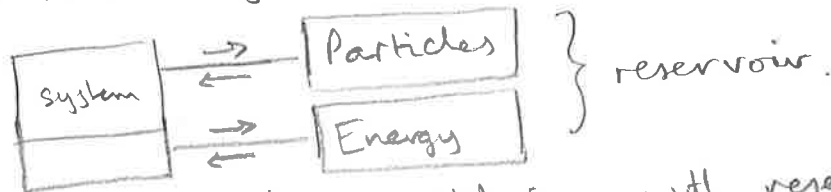
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- A. Affinity chromatography.
 - B. Dialysis to equilibrium (open bag and assay)
 - C. Surface plasmon resonance (for immob. receptors)
change of mass detected by reflectivity.
 - D. Calorimetry

7. The Gibbs Distribution ³⁻⁶ (p289, ch7).

New concept of particle reservoir.

as well as temperature reservoir.

New thermodynamic variable = N = particle number.



System comes to equilibrium with reservoir.

$$N_{tot} = N_s + N_r \quad \text{total particles}$$

$$E_{tot} = E_s + E_r \quad \text{total energy}$$

s = system

r = reservoir

We want the probability:

$P(\text{state})$ \propto no. of states available to reservoir for system in that state.

$$\propto e^{S/k_B} \quad \text{since } S = k_B \ln W_{res}.$$

$$S = k_B \ln W$$

Comparing two states

$$\frac{P(E_s^{(1)}, N_s^{(1)})}{P(E_s^{(2)}, N_s^{(2)})} = \frac{W_{res}^{(1)}}{W_{res}^{(2)}} = \frac{e^{S_r(E_{tot}-E_s^{(1)}, N_{tot}-N_s^{(1)})/k_B}}{e^{S_r(E_{tot}-E_s^{(2)}, N_{tot}-N_s^{(2)})/k_B}}$$

linear expansion of entropy (Taylor series)

$$S_{res}(E_{tot}-E_s, N_{tot}-N_s) = S_{res}(E_{tot}, N_{tot}) - \frac{\partial S_{res}}{\partial E} E_s - \frac{\partial S_{res}}{\partial N} N_s$$

Thermo definitions

$$\Rightarrow \frac{P(E_s^{(1)}, N_s^{(1)})}{P(E_s^{(2)}, N_s^{(2)})} = \frac{e^{-(E_s^{(1)} - \mu N_s^{(1)})/k_B T}}{e^{-(E_s^{(2)} - \mu N_s^{(2)})/k_B T}}$$

Analogous to Boltzmann, all possible microstates, i :

$$P(E_s^{(i)}, N_s^{(i)}) = \frac{1}{Z} e^{-(E_s^{(i)} - \mu N_s^{(i)})/k_B T}$$

Gibbs Distribution

$$Z = \sum_i e^{-(E_s^{(i)} - \mu N_s^{(i)})/k_B T} \quad \text{Grand Partition Function}$$

The Grand Partition Function allows us to obtain the average particle number:

$$\begin{aligned} \langle N \rangle &= \frac{1}{\beta} \frac{\partial}{\partial \mu} \ln Z = \frac{1}{\beta} \frac{\partial}{\partial \mu} \ln \sum_i e^{-\beta(E^{(i)} - \mu N^{(i)})} \\ &= \frac{1}{Z} \frac{1}{\beta} \sum_i \beta N^{(i)} e^{-\beta(E^{(i)} - \mu N^{(i)})} \\ &= \frac{1}{Z} \sum_i N^{(i)} e^{-\beta(E^{(i)} - \mu N^{(i)})} \\ &= \sum_i N^{(i)} p(E^{(i)}, N^{(i)}) \quad \text{usual form.} \end{aligned}$$

This is in addition to $\langle E \rangle = -\frac{\partial}{\partial \beta} \ln Z = -\frac{1}{Z} \frac{\partial}{\partial \beta} Z$.

8. Ligand-Receptor (take 3).

The average $\langle N \rangle$ is the same thing as P_{bound} for a ligand-receptor system. Think of solution as the reservoir.

Each receptor can be in one of 2 states:

$$\left. \begin{array}{l} \sigma = 0 \quad \text{unbound} \\ \sigma = 1 \quad \text{bound} \end{array} \right\} E_{\text{state}} = \epsilon_b \sigma \quad N_{\text{state}} = \sigma$$

$$Z = \sum_{\sigma=0}^1 e^{-\beta(\epsilon_b \sigma - \mu \sigma)} = 1 + e^{-\beta(\epsilon_b - \mu)}$$

$$P_{\text{bound}} = \langle N \rangle = \frac{e^{-\beta(\epsilon_b - \mu)}}{1 + e^{-\beta(\epsilon_b - \mu)}} \quad \text{as before.}$$

Recall connection between μ and concentration:

$$\mu = \mu_0 + k_B T \ln c/c_0$$

$$\Rightarrow \langle N \rangle = \frac{c/c_0 e^{-\beta(\epsilon_b - \mu_0)}}{1 + c/c_0 e^{-\beta(\epsilon_b - \mu_0)}} \quad \text{as seen before}$$

9. Phosphorylation p293

This is used in energy activation to drive processes that are otherwise unfavorable.

Also used as a switch:

Regulation of protein (enzyme) activity.

- much faster than synthesising / denaturing.
- + more efficient for cell.

2-state system:

"Covalent post-translational modification"

kinase / phosphatase on/off or off/on.

OH group of Serine (S)

threonine (T) extra CH_3 .

tyrosine (Y) extra 

in prokaryotes, also histidine (H)

aspartate (D).

10. Allosteric enzyme. (eg Phosphofruktokinase).

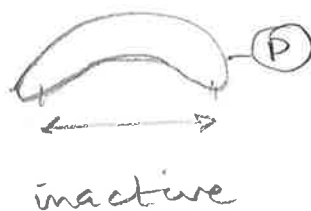
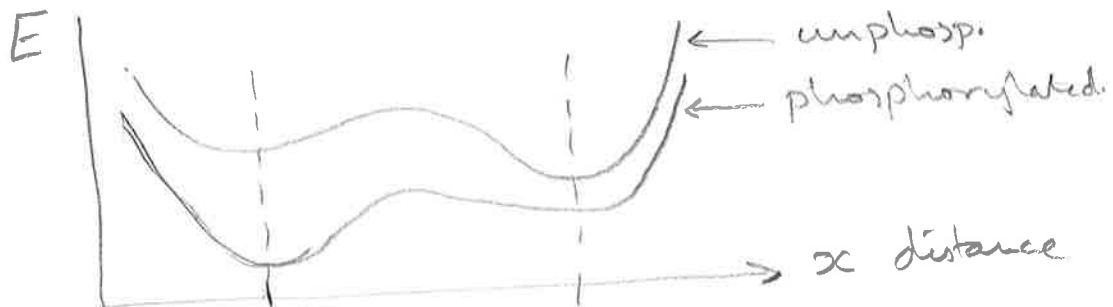
Model of a 2-state protein system.

Catalytic site +

Binding site for control molecule (eg P).

Mechanism of control is to tip the energy landscape: Rapid response.

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might be two folded domains connected by "hinge"

11. Signal transduction p296.

(mostly prokaryotes)

eg lac repressor, switches enzymatic pathway for new food source, lactose.

But food is outside cell,

enzymes are inside, control of DNA.

Signal has to cross cell wall/membrane.

"Sensor histidine kinase" = relay.

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i) binds environmental signal.

ii) enables ATP → Phosphorylation inside

iii) transfers P to "response regulator" aspartate

iv) = "transcription factor" 1000x binds DNA.

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Large variety of responses | eg lac repressor

eg temperature

osmolarity

metal ions

slideHigh specificity demonstrated by
Comparing resp. regulators on gel. (^{32}P)
 ^{31}P is only stable isotope 14-day half-life

12. Hemoglobin.

Cooperativity = binding energy depends number of ligands already bound.

Variant of allostery - mediated by conformation change of protein.

Specifically, O_2 binds to heme which changes shape:

oxy: planar heme config.

deoxy: Fe moves out of plane proximal histidine follows. conformation change.

Electronic state of Fe still not understood.

Fe^{3+} ion smaller than Fe^{2+}

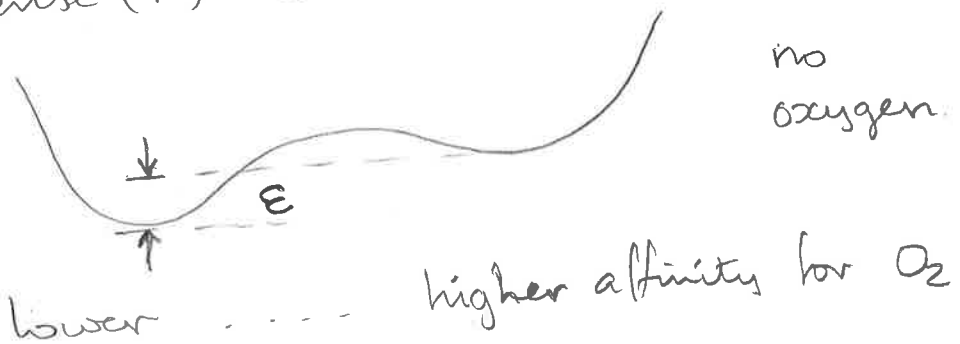
oxy

deoxy

[but Fe in Hb is always diamagnetic]
So no unpaired electrons.

Studied in 1965 by Monod-Wyman-Changeux (MWC).

Tense (T) \rightleftharpoons Relaxed (R).



Binding energy $E_T > E_R$.

Coupling between the two binding energies and the conformation leads to cooperativity.

slides
movies

13 "Dinoglobin"

Simplify to two site MWC model.

Three state variables.

$$\left. \begin{array}{l} \sigma_1 = 0, 1 \\ \sigma_2 = 0, 1 \end{array} \right\} \text{occupation of hemis} \text{ ligand.}$$

$$\sigma_m = 0, 1 \quad \text{T or R state of receptor.}$$

Eight terms in partition function:

$$E = (1 - \sigma_m)(\sigma_1 + \sigma_2)\epsilon_T + \sigma_m[\epsilon + (\sigma_1 + \sigma_2)\epsilon_R]$$

Introduce concentration variables:

$$x = e^{-\beta(\epsilon_T - \mu)} = c/c_0 e^{-\beta(\epsilon_T - \mu_0)} \quad \text{as before}$$

$$y = e^{-\beta(\epsilon_R - \mu)} = e^{-\beta(\epsilon_R - \mu_0)}$$

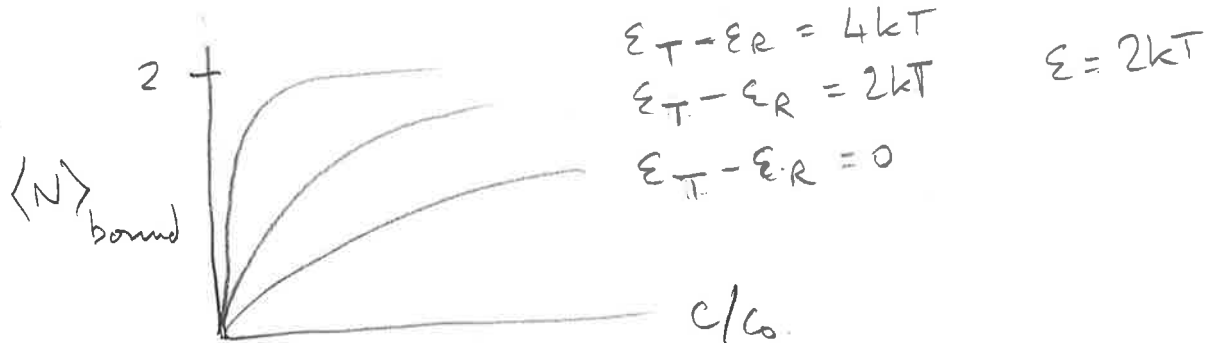
$$\begin{aligned} Z &= 1 + 2x + x^2 + e^{-\beta\epsilon} (1 + 2y + y^2) \\ &= (1+x)^2 + e^{-\beta\epsilon} (1+y)^2 \end{aligned}$$

$$\langle N \rangle = \frac{1}{\beta} \frac{\partial}{\partial \mu} \ln Z$$

$$= \frac{1}{\beta} \frac{1}{Z} (2(1+x)\beta x + e^{-\beta\epsilon} 2(1+y)\beta y)$$

$$= \frac{2}{Z} (x + x^2 + e^{-\beta\epsilon} (y + y^2))$$

Slide



This illustrates the sharper cooperative binding of the MWC model.

14 Ising model.

This is the basis of the Adair and Pauling models used to explain hemoglobin data in 1925.

Interaction between sites is included by a cross-term with its own energy J .

Dimeroglobin again for simplicity

$$\left. \begin{array}{l} \sigma_1 = 0, 1 \\ \sigma_2 = 0, 1 \end{array} \right\} E = \varepsilon(\sigma_1 + \sigma_2) + J\sigma_1\sigma_2 \quad (\text{nonlinear now})$$

Single concentration variable:

$$x = e^{-\beta(\varepsilon - \mu)} = c/c_0 e^{-\beta(\varepsilon - \mu_0)}$$

$$Z = 1 + 2x + x^2 e^{-\beta J}$$

$$\langle N \rangle = \frac{1}{\beta} \frac{\partial}{\partial \mu} \ln Z$$

$$= \frac{1}{\beta} \frac{1}{Z} (2\beta x + 2x\beta x e^{-\beta J})$$

$$= \frac{2}{Z} (x + x^2 e^{-\beta J})$$

Verify correct behaviour when $J \rightarrow 0$

$$\langle N \rangle = \frac{2(x + x^2)}{(1+x)^2} = 2 \frac{x}{1+x} = \text{twice standard L-R result.}$$

When J is large and negative, the x^2 term dominates:

$$\langle N \rangle \rightarrow \frac{2x^2 e^{-\beta J}}{1 + x^2 e^{-\beta J}} = n=2 \text{ Hill function}$$

Binding curve sharpens.

Full model of 4 sites worked in text.

Good agreement with real data of Imai.

Pauling model = single J

Adair model = higher order site interactions.

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